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Dose-dependent Modulation of Lipid Parameters, Cytokines and RNA by δ -tocotrienol in Hypercholesterolemic Subjects Restricted to **AHA Step-1 Diet**

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Authors' contributions

The present study was carried out in collaboration between all authors. Author AAQ was responsible for research planning, analysis of cytokines/miRNA, interpretation and writing the manuscript for publication. Author DAK carried out the study/performed lipids analysis, compilation of study results, the statistical analyses, and revision of manuscript. Author WM did screening/clinical examination of the hypercholestrolemic subjects, data collection and follow up of the subjects. Author NQ critically reviewed protocol and manuscript for publication.

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ABSTRACT

Aims: Evaluate the consumption of δ -tocotrienol (free from tocopherols) on serum lipid parameters, and several cytokines (TNF- α , IL-4, IL-6, IL-8, IL-10), including gene expression and circulating microRNAs (miRNAs) in hypercholesterolemic subjects.

Study Design: The present preliminary dose-response study consisted of six phases. All hypercholesterolemic subjects took increasing doses of δ -tocotrienol (125, 250, 500,750 mg/d) plus AHA Step-1 diet for 4 weeks during the 30 weeks study period.

Methodology: Hypercholesterolemic (n = 31; serum cholesterol > 5.2 mmol/L) subjects (males-26/females 5; age range 50-71 years) were enrolled in the study from Wah Cantonment, Pakistan. Serum lipid parameters were measured by auto analyzers. Various plasma cytokines, cDNA, and miRNAs were estimated by Signosis kits.

Results: All participants (n = 31) completed all phases of study. The δ -tocotrienol plus AHA Step-1 diet caused reductions in lipid parameters in a dose-dependent manner with maximum effects on serum total cholesterol (15%), LDL-cholesterol (18%), triglycerides (14%) with 250 mg/d dose (P< 0.001). Doses above 500 mg/d resulted ininduction in levels of all lipid parameters, except HDL-cholesterol. The cytokines associated with cardiovascular disease (plasma TNF- α , IL-2, IL-4, IL-6, IL-8, IL-10) were all down-regulated 39%-64% by δ -tocotrienol treatment (P< 0.01). Similar results were obtained with gene expression of these cytokines using whole blood messenger-RNA. In contrast, circulating miRNA-7a, miRNA-15a, miRNA-20a (anti-angiogenic), miRNA-21, miRNA-29a, miRNA-92a, miRNA-200, miRNA-206 (skeletal muscle regeneration) down-regulated in hypercholesterolemic subjects, were up-regulated by δ -tocotrienol treatment as compared to baseline (P< 0.01).

Conclusion: The present results confirm that consumption of δ -tocotrienol plus AHA Step-1 diet causes significant reduction in serum lipid parameters and several cytokines (TNF- α , IL-2, IL-4, IL-6, IL-8, IL-10) at a low optimal dose (250 mg/d). The capacity of δ -tocotrienol to modulate inflammation is partly attributable to dose-dependent properties of inhibition/activation, which may play a major role in future treatment of cardiovascular diseases.

Keywords: DeltaGold-90% δ-tocotrienol + 10% γ -tocotrienol; lipid parameters; inflammatory biomarkers; cytokines; TNF- α ; gene expression; circulatory miRNAs.

ABBREVIATIONS

Palm oil TRF: Palm oil tocotrienol rich fraction (Mixture of α -tocopherol, 33.3% + α -tocotrienol,14.5 + γ -tocotrienol, 35.6 + δ -tocotrienol, 16.6%); DeltaGold : 90% δ -tocotrienol + 10% γ -tocotrienol; AHA Step-1 diet: American Heart Association Step-1 diet; HDL: high density lipoprotein; LDL: low density lipoprotein; HMG-CoA reductase: β -hydroxy- β -methylglutaryl-coenzyme A reductase; TNF- α : tumor necrosis factor-alpha; IL: interleukin; mRNA: messenger-ribonucleic acid; miRNA: micro-ribonucleic acid.

1. INTRODUCTION

We have been studying lipid lowering effects of naturally-occurring compounds for several years, such as tocotrienols isolated from palm oil known as tocotrienol rich fraction (TRF), and its components, $\alpha\text{-tocopherol},$ $\alpha\text{-tocotrienol},$ $\gamma\text{-tocotrienol},$ and $\delta\text{-tocotrienol}$ in chickens and humans [1,2]. The tocotrienol rich fraction (TRF) from palm oil, comprising of a mixture of tocopherols and tocotrienols, has shown both positive [3-12] and negative [13-17] hypocholesterolemic effects in a number of

reported clinical studies [2-17]. Palm TRF (palmvitee capsule, 200 mg/day) or rice bran preparation TRF₂₅ low in α -tocopherol concentration (< 10%) combined with AHA Step-1 diet have been effective in lowering serum total cholesterol, LDL-cholesterol, and triglycerides levels in hypercholesterolemic human subjects [2,8]. A major factor underlying failure of other studies to exhibit beneficial effects is attributable to presence of over 20% α -tocopherol in palm TRF. This probably inhibited TRF from lowering serum total cholesterol or LDL-cholesterol levels in four major studies [14-17]. Palm TRF also

does not reduce serum total cholesterol level in free-living hypercholesterolemic patients [15-17], or healthy humans even if the TRF contained less than 15% α -tocopherol [13]. Furthermore, large doses of tocotrienols have also proved ineffective [8,18-20] perhaps owing bioconversion of tocotrienols to α -tocopherol, which antagonizes this beneficial effect [8]. The serum level of α -tocopherol was 2 to 4 fold higher, as compared to the placebo group in these studies [14-17]. Therefore, high dose of tocotrienols are not very effective in reducing serum levels of total cholesterol, LDL-cholesterol, and triglycerides.

We accordingly carried out a study with pure tocotrienols devoid of tocopherols, instead of TRF from palm oil which contains variable concentrations of α -tocopherol. The availability of tocopherol-free DeltaGold from annatto seeds (consisting of 90% δ -tocotrienol + 10% γ tocotrienol; Fig. 1) made this human study possible. We have previously demonstrated the mechanism underlying through tocotrienols exert their effects by suppressing the activation of nuclear factor-κB (NF-κB) in various experimental models [21]. Moreover, the order of potency of various tocotrienols for acting as cholesterol-lowering, anti-oxidant. antiinflammatory and anticancer agents were as follows: δ -tocotrienol > γ -tocotrienol > α tocotrienol > α -tocopherol [22,23]. Recently, a comprehensive review has compared the various biological properties of tocotrienols, including results of various clinical studies of palm TRF and pure tocotrienols [24].

Aside from measuring lipid parameters (total cholesterol and LDL-cholesterol) as classic indicators of cardiovascular disease risk factors, the present study also examined inflammatory cytokines implicated in heart disease and their

gene expression. These included tumor necrosis factor-alpha (TNF- α), a cytokine which is an important contributor to atherosclerotic lesion development [25], interleukin-2 (IL-2) level of which is significantly elevated in patients with stable angina [26], interleukin-4 (IL-4), an activator of collagen synthesis that may be involved in cardiac fibrosis [27], interleukin-6 (IL-6) continuous production of which promotes production myocardial injury and can cause cardiac hypertrophy [28], and interleukin-8 (IL-8), a cytokine found in vascular injury sites that plays a role in various stages of atherosclerosis [29].

The expression of circulating microRNAs (miRNA) which are small non-coding RNAs, that are likely involved in many biological processes were also analyzed [30,31]. The present study evaluated effect of δ -tocotrienol on selected miRNAs associated with cardiovascular disease such as miRNA-7a, miRNA-15a, miRNA-20a, miRNA-21, miRNA-29a, miRNA-92a, miRNA-200, and miRNA-206. Particularly, miRNA-29a was examined, a family that accounts for ~4% of all miRNAs in the murine heart [30]. MicroRNA-29a is down-regulated after myocardial infarction (MI), targets genes involved in fibrosis and is known as a fibrotic inhibitor. Other miRNAs examined in the present study included antiangiogenic miRNA-20a, and miRNA-206, which mainly promotes skeletal muscle regeneration, but may also play a pivotal role in the heart muscle [31]. The present study of dose-response (125, 250, 500,750 mg/d) of feeding DeltaGold (90% δ-tocotrienol + 10% γ-tocotrienol) plus AHA Step-1 diet tohypercholesterolemic subjects was carried out on serum lipid parameters, various plasma cytokine levels, and their gene expression and plasma circulating miRNA levels associated with cardiovascular disease.

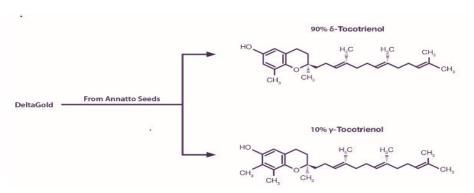


Fig. 1. Chemical structures of DeltaGold (90%δ-tocotrienol + 10% γ-tocotrienol)

2. MATERIALS AND METHODS

The study was carried out in the Department of Chemical Pathology & Endocrinology, Armed Forces Institute of Pathology (AFIP), Rawalpindi, Pakistan in collaboration with the Department of Basic Medical Sciences, University of Missouri-Kansas City, MO, USA The study protocol was registered and approved by Institutional Review Board of AFIP, Rawalpindi, Pakistan. The study was carried out under a FDA approved IND number 36906.

2.1 Materials

DeltaGold 125 mg soft gels from annatto seeds (composition 90% δ-tocotrienol + 10% γ-tocotrienol) were supplied by American River Nutrition, Inc. (Hadley, MA. USA). Serum total cholesterol, HDL-cholesterol, LDL-cholesterol, and triglycerides levels were estimated by using reagent kits from Sigma Chemical Co., St. Louis. Pure total RNA was obtained from the EDTA treated fresh whole blood by using" total RNA purification kit # 17200 (NORGEN Bioteck Corporation, Thorold, ON, Canada). The various plasma cytokines, cDNA, and miRNA were estimated by using Signosis, Inc. (1700 Wyatt Drive Suite 10-12. Santa Clara, CA). Human Elisa Plate Arrav (chemiluminescence), Catalog number EA-4001, Customized Human cDNA Plate Array (Catalog Number AP-UM000416) from messenger ribonucleic acid (mRNA). The mRNA was extracted from each sample and converted to cDNA and plated on a cytokine cDNA array plate (Signosis, Inc.). Estimation of circulating microRNAs (miRNAs) was carried out using customized MiRNA Direct Hybridization Plate Array (chemiluminescence; Catalog Number Inv-00465) according to the manufacturer's instructions (Signosis, Inc.).

2.2 Study Design

The present study was a forced titration design, where all subjects took increasing doses (125, 250, 500, 750 mg/d) of δ -tocotrienol plus AHA Step-1 diet after baseline (phase I) and AHA Step-1diet (phase II). A sample size of this study (n=31) was based on data derived from senior citizens with alpha 0.05 and beta 0.8 to assess the effectiveness of the tocotrienols in different doses (Mammatech Inc., Coppell, Texas, USA).

The study subjects were screened for high cholesterol from the general community at Wah Cantonment, Pakistan. Clinical history was taken and physical examination was carried out for each participant. The initial measures included the participant's height, weight, systolic and diastolic blood pressure at rest, history of significant diseases, medications (including statins, nitrates, calcium antagonists, angiotensin-converting enzyme [ACE] inhibitors, and/or diuretics) and tobacco smoking. The height and weight were measured in light clothing and without shoes. Body mass index (BMI, kg/m²) was calculated for each subject.

2.2.1 The inclusion criteria

Adults male/female, age > 50 years with cholesterol level ≥ 5.2 mmol/L labeled as hypercholesterolemic were included in the study [32].

2.2.2 The exclusion criteria

Any subject having weight (> 125% of Metropolitan Life relative weights), taking cholesterol lowering medication or anti-inflammatory drugs in the last 2 weeks were excluded. The subjects with elevated serum transaminase activity, serum urea, glucose, thyroid stimulating hormone, liver, renal, diabetes, and thyroid diseases were excluded from the study. A total of (n = 31) hypercholesterolemic subjects (26 males + 5 females) were enrolled in this study.

All subjects signed an informed-consent form, which was approved by the Institutional Review Board of Armed Forces Institute of Pathology, Rawalpindi, Pakistan. Each participant was individually counseled to American Heart Association (AHA) Step-1 diet (restricted intake of fat < 30% / d, and cholesterol < 300 mg/d) throughout the study period. Participants of the study were also advised to stop using cholesterol-lowering drugs or anti-oxidants and counseled individually to modify food intake to meet the goals of the AHA Step-1 diet. Subjects were asked to stop the intake of whole milk, butter, cheese, eggs, animal fat and ice cream. In order to ascertain full adherence to dietary recommendations and intake of nutritional supplements, participants were contacted by telephone during each phase.

2.3 Experimental Design

2.3.1 Effect of δ-tocotrienol plus AHA Step-1 diet in hypercholesterolemic subjects

The experiment consisted of six phases; the first (phase I), an alcohol-free choice diet phase (baseline) was followed by a 4 week second phase (phase II), during which all participants were counseled to follow the American Heart Association Step-1 diet (AHA Step-1diet). All participants were continued on the AHA Step-1 diet during phases III, IV, V and VI. During phase III, all participants were administered 1 capsule (125 mg/d) of δ -tocotrienol (8 pm after food) for 4 weeks. During phase IV, participants were administered 2 capsules of 125 mg (250 mg/d; one at 8 am and one at 8 pm after breakfast and dinner) for 4 weeks, followed by 4 capsules of 125 mg (500 mg/d; two at 8 am and two at 8 pm) in phase V and during the last phase VI, 6 capsules of 125 mg (750 mg/d; two at 8 am, two at 2 pm and two at 8 pm after food) were administered for 4 weeks as outlined in Fig. 2. There was a 2 week washout period after the treatment of the first dose of 125 mg/d, however. all subjects were continued on AHA Step-1 diet for the rest of the treatment period. At the end of each phase, blood samples were collected after overnight fast of each participant to carry out estimation of lipid parameters and several inflammatory biomarkers. Serum/plasma samples from all the subjects of each group were studied simultaneously to avoid large standard variation/deviation.

2.4 Blood Collection

Venous blood samples (12 h fast, 9:00 pm – 9:00 am) were drawn at screening. At screening, the

participants were counseled to follow their normal dietary intake. Screening was accomplished during three to four weeks (baseline). Venous blood samples were drawn at the termination of baseline phase, and at week four of the treatment. The processed samples were coded and held at -72°C until analyses were carried out, following the completion of treatment phases.

2.5 Analyses

The analyses of the coded samples were performed at the department of Chemical Pathology and Endocrinology, Armed Forces Institute of Pathology, Rawalpindi, Pakistan. The analyses of samples of all the phases for each parameter were carried out at the same time to avoid large variation. Serum total cholesterol, HDL-cholesterol, LDL-cholesterol, triglycerides levels were measured in each sample for every subject. Automated clinical laboratory procedures were used for determining lipid parameters at the end of phase I (4 weeks); II (8 weeks); III (12 weeks), IV (18 weeks), V (24 weeks), and VI (30 weeks). Serum LDLcholesterol levels were estimated by precipitating 200 µL of serum with 25 µL of a mixture of 9.7 mM phosphotungstic acid and 0.4 M MgCl₂. The preparation was mixed for 10 min at room temperature and then centrifuged at 12,000 x g for 10 min. The supernatant fraction was decanted and analyzed for levels HDL-cholesterol. The precipitate was dissolved in 200 µL of 0.1 M sodium citrate and LDLcholesterol level was determined [33]. Serum total cholesterol. HDL-cholesterol. LDLcholesterol, and triglycerides levels were estimated by using reagent kits (Sigma Chemical Co., St. Louis).

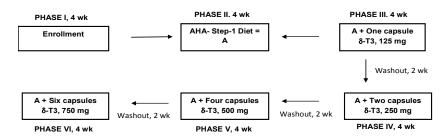


Fig. 2. Annatto-based δ -tocotrienol treatment study protocol corresponds to six phases, and each phase lasted for 4 weeks

Enrollment (baseline) = I; AHA Step-1 diet = II; δ -tocotrienol 125 mg/d + AHA Step-1 diet = II; δ -tocotrienol 250 mg/d + AHA Step-1 diet = IV; δ -tocotrienol-500 mg/d + AHA Step-1 diet = V; δ -tocotrienol 750 mg/d + AHA step-1 diet = V1, fed to hypercholesterolemic subjects.

2.6 Analyses of Total RNA from EDTA Treated Whole Blood after Feeding δ-tocotrienol Plus AHA Step-1 Diet for 4 Weeks to Hypercholesterolemic Subjects

The pure total RNAs were extracted from EDTA treated fresh whole blood drawn from subjects those were fed the most effective dose of δ tocotrienol (250 mg/d) plus AHA Step-1 diet for 4 weeks, and total RNA purification kit # 17200 (NORGEN Bioteck Corporation, Thorold, ON, Canada) was used for this purpose. The purity of total RNA was carried out by measuring the absorption at several wavelengths using a Scientific NanoDrop 1000 Thermo Spectrophotometer. The purity of total RNA was determined by the ratio of 260/280 (2.02 - 2.08). The plasma miRNAs (dose of 250 mg/d of δ tocotrienol plus AHA Step-1diet fed for four weeks) were also purified bγ Plasma/Serum Circulating miRNA Purification Mini Kit (Slurry Format) Product # 51000 (NORGEN Bioteck Corporation, Thorold, ON, Canada).

2.7 Estimation of Human Plasma Cytokines, cDNA, and miRNA

The various plasma cytokines, cDNA, and microRNAs (miRNAs) were estimated by using Human Cytokine Elisa Plate Array (chemiluminescence), Catalog number EA-4001, Customized Human cDNA Plate Array (Catalog Number AP-UM000416) from messenger-RNA (mRNA) (Signosis, Inc., Santa Clara, CA, 95054). The mRNA extracted from each sample was converted to cDNA and plated on a cytokine cDNA array plate (Signosis, Inc.). Assays for estimating the plasma cytokines (protein) and gene expression of messenger RNAs were carried out according to the protocols provided by Signosis, Inc. The incubation of each assay mixture at various temperatures was carried out by using Enviro-Genie Shaker/incubator (Enviro-Genie Industries. Bohemia. NY). The intensity of chemiluminescence was detected using a Microplate Luminometer (GloMax Promega, Madison, WI) at 500 nm, and luminescence was monitored over 20 min period. Estimation of circulating miRNAs was carried out using "Customized miRNA Direct Hybridization Plate Array", chemiluminescence; Catalog Number Inv-00465 (Signosis, Inc).

2.8 Statistical Analyses

The data were analyzed by using the GLM procedure of SAS (Statistical Analysis System) for personal computers to test the study hypothesis. Analysis of two-way variance was used to test whether changes in serum lipid parameters occur during the course of supplementation, and whether there were within-subject betweenand differences; because all observations were required, available degree of freedom were reduced by this statistical approach [34]. Data are reported as mean ± SD (Standard Deviation). The statistical significance level was set at P< 0.05.

3. RESULTS

3.1 Inhibitory Effects of δ-tocotrienol plus AHA Step-1 Diet on Lipid Parameters in Hypercholesterolemic Subjects

The commercial availability of DeltaGold (90% δtocotrienol + 10% γ-tocotrienol) from annatto seeds enabled us to carry out dose-response study of 125 mg, 250 mg, 500 mg and 750 mg/d plus AHA Step-1 diet (restricted intake of fat < 30%/d, and cholesterol < 300 mg/d) in hypercholesterolemic subjects. All participants (n = 31) completed all phases of the study, and there was no change in the body weight, and other physical characteristics of the participants during the treatment period (Table 1). There were insignificant reductions of 2%, 3%, 3% in serum levels of total cholesterol, LDL-cholesterol and triglycerides, respectively, due to dietary restriction (AHA Step-1 diet) after 4 weeks, as compared to baseline values (Figs. 3-5). However, consumption of δ -tocotrienol plus AHA Step-1 diet lowered serum total cholesterol, LDLcholesterol and triglycerides levels in a dosedependent manner below 500 mg/d, in contrast, higher dose of 750 mg/d increased levels of these lipid parameters (Figs. 3-5).

The optimal dose was found to be 250 mg/d of δ -tocotrienol, plus AHA Step-1 diet which after 4 weeks caused significant reductions of serum total cholesterol (15%; P < 0.001), LDL-cholesterol (18%; P < 0.001) and triglycerides (14%; P < 0.001) levels, compared to baseline (Figs. 3, 4, 5). The administration of minimum dose of 125 mg/d of δ -tocotrienol plus AHA Step-1 diet did not cause any remarkable reductions in serum levels of total cholesterol, LDL-cholesterol

and triglycerides (3%, 6%, 2%), respectively, compared to baseline (Figs. 3-5). This slight reduction might be due to AHA Step-1 diet restriction. Administration of highest dose (750 mg/d) of δ-tocotrienol plus AHA Step-1 diet after 4 weeks resulted inincreases of 10%, 15%, 9% in serum levels of total cholesterol, LDL-cholesterol, and triglycerides respectively, compared to a dose of 250 mg/d + AHA Step-1 diet, which might be due to novel properties of δ -tocotrienol (Figs. 3, 4, 5). Serum HDL-cholesterol level was not affected compared to baseline under these conditions (data not shown). Similar trends of increases or decreases in levels of total cholesterol, LDL-cholesterol, triglycerides, and HDL-cholesterol with various doses also impact the ratios of total cholesterol/HDL-cholesterol LDL-cholesterol/HDL-cholesterol compared to baseline (data not shown). The efficacy and safety assessment was done at the end of each phase of increasing doses of tocotrienol treatment. The efficacy was analyzed based on the changes in the lipid parameters as compared with baseline levels. Regarding safety and tolerability of different doses of tocotrienols, hypercholesterolemic subject did not report any adverse events during the study, and there was no adverse effect or reaction after use of the higher dose of 750 mg/d or the minimum dose of 125 mg/d of δ -tocotrienol reported by any participant throughout the treatment period of 4 weeks each. Therefore, administration of 125 – 750 mg/d of DeltaGold (90% δ -tocotrienol + 10% γ -tocotrienol) was tolerable and safe for human consumption. Coincidently, GeltaGold has been granted "GRAS" status by FDA recently [GRAS (Generally Regarded As Safe) Notice No: GRN 000471].

Table 1. Baseline characteristics of study population

Parameters	Means ± SD
Age (years)	57.84 ± 8.07
Male/Female(n)	26/5
Height (meter)	1.74 ± 0.07
Weight (Kg)	69 ± 7
BMI (Kg/m ²)	25.30 ± 1.86
Systolic BP (mmHg)	140.16 ± 6.26
Diastolic BP (mmHg)	90.32 ± 5.31
Blood glucose (mmol/L)	4.22 ± 0.43
Serum creatinine (µmol/L)	93.39 ± 10.12
Serum ALT (U/L)	36.68 ± 7.97
Serum cholesterol (mmol/L)	5.44 ± 1.06
Serum triglycerides (mmol/L)	1.81 ± 0.54

The treatments 1- 6 correspond to six phases as shown in Fig 2. Data are means \pm SD (Standard Deviation). Values in a column not sharing a common symbol are significantly different at $\P = P < 0.001$; $\S = P < 0.05$.

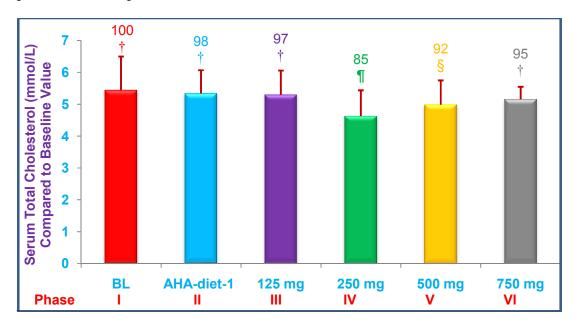


Fig. 3. Inhibitory effects of various doses of δ -tocotrienol plus AHA Step-1 diet on serum levels of total cholesterol in hypercholesterolemic subjects

The treatments 1- 6 correspond to six phases as shown in Fig. 2. Data are means \pm SD (Standard Deviation). Values in a column not sharing a common symbol are significantly different at $\P = P < 0.001$; $\S = P < 0.05$.

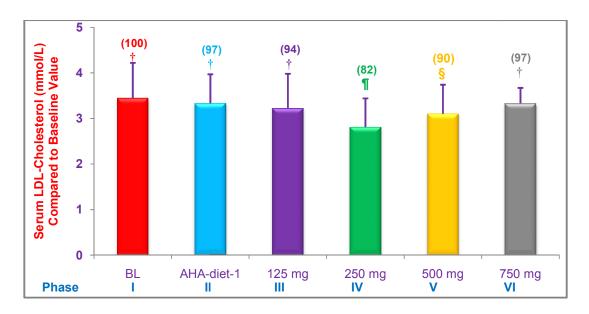


Fig. 4. Inhibitory effects of various doses of δ-tocotrienol plus AHA Step-1 diet on serum levels of LDL-cholesterol in hypercholesterolemic subjects

The treatments 1- 6 correspond to six phases as shown in Fig. 2. Data are means \pm SD (Standard Deviation). Values in a column not sharing a common symbol are significantly different at $\P = P < 0.001$; $\S = P < 0.03$.

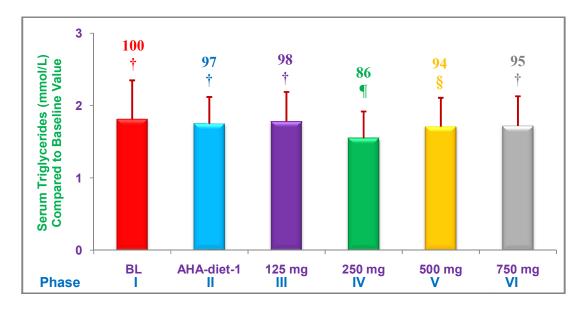


Fig. 5. Inhibitory effects of various doses of δ -tocotrienol plus AHA Step-1 diet on serum levels of triglycerides in hypercholesterolemic subjects

The treatments 1- 6 correspond to six phases as shown in Fig. 2. Data are means \pm SD (Standard Deviation). Values in a column not sharing a common symbol are significantly different at $\P = P < 0.001$; $\S = P < 0.05$.

3.2 Evaluation of Feeding δ-tocotrienol Plus AHA Step-1 Dieton Levels of Cytokines, Gene Expression and miRNA in Hypercholesterolemic Subjects

A panel of six key plasma cytokines associated with cardiovascular disease (TNF-α, IL-2, IL-4, IL-6, IL-8, IL-10) was selected to investigate the anti-inflammatory and cardio-protective effect of δ-tocotrienol taken orally. The functions of each of these cytokines are reported in Table 2. The AHA Step-1 diet alone did not have any significant effect on the levels of plasma cytokines except on IL-8 (Table 2). However, the treatment with δ-tocotrienol plus AHA Step-I diet showed down-regulated levels of TNF-α. IL-2. IL-IL-8, and IL-10 (39% - 64%) as compared to baseline values. The maximum reduction was observed in IL-6 cytokine, which acts as both a pro-inflammatory and antiinflammatory cytokine, and is secreted by T-cells and macrophages to modulate immune response (Table 2). The present results show downregulation of IL-6 and IL-8 levels by δ -tocotrienol, confirming the anti-angiogenic properties of δ tocotrienol. These cytokine data can be very well correlated to gene expression of messenger-RNA (mRNA) purified from fresh EDTA treated whole blood obtained from subjects on the same treatment (250 mg/d; Table 3).

The cluster of eight microRNAs (miRNA-7a, miRNA-15a, miRNA-20a, miRNA-21, miRNA-29a, miRNA-200, miRNA) was typically down-regulated in hypercholesterolemic subjects (baseline values) as shown in Table 4.

The δ -tocotrienol plus AHA Step-1 diet treatment up-regulated miRNAs as compared to baseline values (Table 4). The AHA Step-1 diet treatment resulted only in slight up-regulation in these miRNAs. These results indicated that δ -tocotrienol treatment up-regulated a cluster of selected miRNAs levels in plasma of hypercholesterolemic subjects.

5. DISCUSSION

The maximum decreases of 2% to 3% in lipid parameters resulted due to AHA Step-1 diet dietary modification in the present study confirming our earlier findings [7-9]. The present results of dose-response study demonstrate that

δ-tocotrienol specifically lowered the levels of serum total cholesterol, LDL-cholesterol, and triglycerides in a dose-dependent manner below 500 mg/d, while at higher dose of 750 mg/d increased levels of these three lipid parameters compared to 250 mg/d + AHA Step-1 diet (Figs. 3-5). These results are consistent with our recent of dose-dependent inhibition chymotrypsin-like activity of 20S rabbit muscle proteasomes between 5 μM and 40 μM for mevinolin and δ -tocotrienol, where the inhibitory effects of mevinolin and δ -tocotrienol were reversed at higher concentrations between 80 μM and 320 μM [21]. This clearly demonstrates that δ-tocotrienol and mevinolin modestly inhibit or activate the proteasomal activity depending on its concentrations [21,35-37]. Thus, δ -tocotrienol is the first naturally-occurring compound, which blocks the proteasomal activity at low doses, and is able to halt and reduce the inflammatory response. This property of δ -tocotrienol may be useful for the control of cardiovascular disease. and at higher doses may cause apoptotic cell death in various types of cancers [38]. Similar dose-dependent activities (inhibition versus induction) and properties have been reported for synthetic proteasomal inhibitors, MG132 and lactacystin [35-37]. The aforementioned are very potent proteasome inhibitors in the range of 5 μ M to 20 μM , but very toxic as well, restricting their use in humans. Conversely, tocotrienols have been found safe even at doses of 1600-3200 mg/d in the treatment of pancreatic cancer [38].

Moreover, a dose of 250 mg/d causes significant reductions in all three lipid risk factors (total cholesterol, LDL-cholesterol, and triglycerides) after 4 weeks of treatment compared to baseline. The lower dose of 125 mg/d may have shown additional lipid lowering benefits in humans, provided the treatment period had been extended to 8 weeks or more. As reported earlier, the hepatic HMG-Co A reductase activity is inhibited by low doses of γ - and δ -tocotrienols, whereas at high doses, tocotrienols may convert to tocopherols, in particular, α -tocopherol which induces the activity of HMG-Co A reductase (a rate-limiting enzyme in the biosynthesis of cholesterol) and consequently raises cholesterol [24,39]. This disadvantage of using high dose of tocotrienols does not apply to their other functions, such as cancer chemoprevention and treatment, where large doses are used in current clinical trials, and may work by activating the apoptosis [38].

Table 2. Evaluation of role of δ-tocotrienol (250 mg/d) + AHA Step-1 diet on various plasma cytokines in hypercholesterolemic subjects

#	Cytokines	Baseline	AHA Step-1 diet = A	A = δ-T3	Description	Functions
	Down-Re	Percentages	Percentages	Percentages		_
1	TNF-α	100	91.0 ± 1.41 ^a	48.5 ± 0.70**	Tumor Necrosis Factor-α	Produced during inflammation.
2	IL-2	100	94.0 ± 1.41	55.5 ± 0.71**	Interleukin-2	for growth proliferation & differentiation of T cells to become "Effect or T cells".
3	IL-4	100	93.0 ± 1.41	49.0 ± 1.41**	Interleukin-4	Activation of B-cells& T cells proliferation.
4	IL-6	100	98.0 ± 1.41	38.5 ± 2.21**	Interleukin-6	Regulates immune response& hematopoiesis.
4	IL-8	100	85.5 ± 2.12*	43.5 ± 0.71**	Interleukin-8	Potent anti-angiogenesis factor.
6	IL-10	100	92.5 ± 2.02	63.5 ± 2.12**	Interleukin-10	Immuno-regulation & inflammation.

 $^{{}^{}a}X \pm SD$ (mean \pm Standard Deviation); δ -T3 = δ -tocotrienol;

Table 3. Evaluation of role of δ-tocotrienol (250 mg/d) + AHA Step-1 diet on gene expression of cytokines in hypercholesterolemic subjects

	Gene Expr	Baseline	AHA Step-1	AHA Step-1	Description	Functions
	Cytokines		diet	diet + δ-T3		
#		Percentages	Percentages	Percentages		
1	TNF-α	100	96.3 ± 2.76 ^a	84.5 ± 0.71 ^{a,**}	Tumor necrosis Factor-α	Inflammation
2	IL-2	100	98.8 ± 0.95	91.5 ± 3.54*	Interleukin-2	Cytokine involved in proliferation, & differentiation.
3	IL-4	100	96.2 ± 0.40	77.5 ± 2.12**	Interleukin-4	Activation of B-cells & T-cells proliferation
4	IL-6	100	95.0 ± 0.86	73.5 ± 0.71**	Interleukin-6	NF-κB and IL-6 signaling.
4	IL-8	100	97.0 ± 1.17	92.0 ± 2.83*	Interleukin-8	Chemokine (involved in angiogenesis).
6	IL-10	100	97.3 ± 0.65	89.0 ± 1.41*	Interleukin-10	Immuno-regulation and inflammation.

 $^{^{}a}X \pm SD$ (mean $\pm Standard Deviation); <math>\delta -T3 = \delta -tocotrienol$.

Table 4. Evaluation of role of δ-tocotrienol (250 mg/d) + AHA Step-1 diet on plasma circulating miRNAs of cardiovascular disease in hypercholesterolemic subjects

	MicroRNA =	Baseline	AHA Step-1 diet	AHA Step-1 diet + δ-T3
	miRNA	Percentages	Percentages	Percentages
1	miRNA-7a	100	103.5 ± 2.12 ^a	168.0 ± 1.41**
2	miRNA-15a	100	107.6 ± 0.71*	179.0 ± 1.41**
3	miRNA-20a	100	102.5 ± 0.71	168.0 ± 2.24**
4	miRNA-21	100	108.0 ± 2.83*	143.0 ± 2.83**
5	miRNA-29a	100	102.5 ± 0.71	142.0 ± 2.83**
6	miRNA-92a	100	106.5 ± 2.12*	153.5 ± 2.12**
7	miRNA-200	100	104.0 ± 1.41	146.0 ± 1.41**
8	miRNA-206	100	109 .0 ± 2.83*	150.0 ± 2.83**

 $^{^{}a}X \pm SD$ (mean \pm Standard Deviation); δ -T3 = δ -tocotrienol.

^{***}Values in a row sharing a common asterisk are significantly different at ***P** < 0.05; ****P** < 0.01.

^{***}Values in a row sharing a common asterisk are significantly different at ***P** < 0.05; ****P** < 0.01.

^{****}Values in a row sharing a common asterisk are significantly different at***P**< 0.05; ****P**< 0.01.

It is also interesting to note that synthetic α tocopherol at 400 IU/day was shown to increase the risk of prostate cancer by 17% in a large "Selenium and Vitamin E Cancer scale Prevention Trial (SELECT)" [40]. It is well documented that high cholesterol is associated with increased risk of prostate cancer [41-44], and prostate cancer cells accumulate cholesterol to spur their growth [45]. Thus it is plausible that the elevated prostate cancer risk of the above study is due to α -tocopherol's stimulation of the cholesterol synthesis pathway [40], while were indicated as potential tocotrienols therapeutic agents for prostate cancer owing to their ability to lower and degrade a major transcription factor in the cholesterol synthesis pathway [45]. Our present study reported no adverse events with large tocotrienol doses, suggesting that δ -tocotrienol at doses as high as 750 mg/day is safe for human consumption. Pure δ-tocotrienol may be safe for consumption even at doses of 3,200 mg/d, as was shown in a recent Phase I Clinical Trial in patients with pancreatic cancer [38].

Recently, inflammation has been shown to be associated with several diseases including cardiovascular disorders [26]. The present study demonstrates that δ-tocotrienol effectively downregulated inflammatory cytokines and gene expression of TNF- α , IL-2, IL-4, IL-6, and IL-8. The maximum down-regulation occurred with IL-6, which is both a pro-inflammatory cytokine (in the case of chronic inflammation oncogenesis) and anti-inflammatory cytokine (in the case of immune regulation and support of hematopoiesis) [46]. While various studies have confirmed tocotrienol's anti-inflammatory functions, particularly for TNF- α and on a proteasomal level [21,23,47], they are also known to support the immune system [48]. Hence they do not appear to adversely affect the anti-inflammatory properties of IL-6. In the present study, results showing down-regulation of IL-6 and IL-8 levels by δ -tocotrienol confirm the anti-angiogenic properties of δ-tocotrienol in pathological conditions. The down-regulation of IL-6 also indicates an effect on NF-κB, by which this cytokine is expressed. Tocotrienol's effect on NF-κB and cytokine expression has been shown earlier [21]. Interleukin-10 (IL-10) is capable of inhibiting several pro-inflammatory cytokines such as TNF- α , IL-2, IFN- γ , and granulocyte macrophage-colony stimulating factor produced by macrophages, regulatory T-cells (Th2), and mast cells, stimulate B cell maturation and

antibody production. IL-10 was modestly increased in premature coronary disease [49].

The levels of miRNA have been shown to be important regulators of gene expression that modify cellular responses and function [50-52]. The dysregulation of miRNA plays a crucial role in the development of cardiovascular disease, diabetes and cancer. In the present study, we focused on miRNA involved cardiovascular disease [52]. The effect of δ tocotrienol's on miRNAs may have important implications in the management of chronic diseases. The present study found that $\delta\text{--}$ tocotrienol up-regulated miRNA-7a, miRNA-15a, miRNA-20a, miRNA-21, miRNA-29a, miRNAmiRNA-200 and miRNA-206 hypercholesterolemic humans. MicroRNAs play multiple roles in various biological processes as well as normal physiological functions, and may also display pathological activity. Since levels of eight miRNAs tested in the present study were down-regulated in the hypercholesterolemic population as compared to normal cholesterolemic subjects according to published report [52], up-regulation by δ tocotrienol of these miRNAs points to a beneficial effect of tocotrienols. MicroRNA-29a is enriched in fibroblasts and encodes proteins involved in fibrosis, including collagen, fibrillins, and elastin [53]. In myocardial infarction and associated cardiac hypertrophy, miRNA-29a is decreased, allowing for expression and deposition of collagen components in the fibrotic scar [30]. Upregulation of miRNA-29a such as with δ tocotrienol may provide a significant therapeutic option for myocardial infarction (MI), reducing scar formation in post-myocardial infarction remodeling.

MicroRNA-20a is anti-angiogenic, and known to inhibit the proliferation and metastasis of pancreatic cancer [54]. It also prevents myocardial hypertrophy and angiogenesis during stress [55]. By up-regulating miRNA-20a, δ tocotrienol may decrease angiogenesis during stress situations to prevent abnormal increase of heart size. Similarly, miRNA-206 that is essential in promoting skeletal muscle regeneration delays the progression of amyotrophic lateral sclerosis [56], while suppressing gastric cancer cell growth and metastasis [57]. While there may be important implications for δ -tocotrienol in these applications [58], the present study focused on the supplement's relevance in cardiovascular diseases. Skeletal muscle degeneration. ameliorated by miRNA-206, was found to

contribute to cardiac dysfunction [59], and hence miRNA-206 may play a pivotal role in the heart muscle [31]. δ -Tocotrienol's up-regulation of miRNA-206 may contribute to myocardial and vascular regeneration, as demonstrated by a previous study in murine chronically failing hearts [60]. The positive impact of γ -tocotrienol of remaining miRNAs has been described in detail in recent publication [61].

It should be pointed out that the present preliminary study started as a double-blind study, but during phase V, most of the participants realized that they were involved in a doseresponse study, however, most effective dose of 250 mg/d of δ -tocotrienol was found to be responsible for lipid-lowering hypercholesterolemic subjects. In order validate the results of the present study on the efficacy of δ-tocotrienol as hypocholesterolemic and anti-inflammatory agent, a larger, more comprehensive double-blind long-term (12 months) study should be carried out by enrolling number of male and hypercholesterolemic subjects (50 of each), a placebo group (corn oil or olive oil stripped of tocols after extraction with absolute ethanol) should be added, and subjects of one more group should be kept on the AHA Step-1 diet for at least 4 months (administered placebo capsule) to establish the long-term impact of dietary restriction in hypercholesterolemic subjects.

6. CONCLUSION

The present results indicate that doses below 500 mg/d of δ-tocotrienol (250 mg/d) administered for 4 weeks are effective in lowering lipid parameters, down-regulating several inflammatory biomarkers (TNF- α , IL-4, IL-6, IL-8, and IL-10) and in contrast, doses above 500 mg/d of δ tocotrienol (750 mg/d) up-regulate biomarkers and possibly kill cancer cells. Therefore, the capacity of tocotrienols to modulate inflammation may be attributable, in part, to their dose-dependent properties of inhibition of gene expression in cardiovascular disease and for activation of apoptosis pathways to kill cancer cells. δ-Tocotrienol was also found to be a potent naturally-occurring compound, which could alter the dysregulation of a number of miRNAs (miRNA-7a, miRNA-15a, miRNA-20a, miRNA-21, miRNA-29a, miRNA-92a, miRNA-200, miRNA-206) levels in hypercholesterolemic subjects. Future investigations may explore the combined therapy of δ -tocotrienol and other naturally-occurring compounds (resveratrol,

quercetin, curcumin) having complementary mechanisms of action as more effective formulation for patients with dyslipidemia, and hypercholesterolemia, and may play a major and significant role in the future management of cardiovascular disease.

CONSENT

All authors declare that they have read the manuscript, and written informed consent was obtained from each subject (or any other approved parties) for publication of this study.

ETHICAL APPROVAL

All authors hereby declare that the trial has been examined and approved by the Independent Ethics Committee of Armed Forces Institute of Pathology (AFIP), Rawalpindi, Pakistan and have been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki.

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COMPETING INTEREST

The authors declare that they have no competing interests.

REFERENCES

 Yu GS, Thomas AM, Gapor A, Tan B, Qureshi N, Qureshi AA. Dose-response impact of various tocotrienols on serum lipid parameters in 5-week-old female chickens. Lipids. 2006;41:453-461.

- Qureshi AA, Bradlow BA, Brace L, Manganello J, Peterson DM, Pearce BC, Wright JJK, Gapor A, Elson CE. Response of hypercholesterolemic subjects to administration of tocotrienols. Lipids. 1995;30:1171-1177.
- Yuen KH, Wong JW, Lim AB, NG BH, Choy WP. Effects of mixed-tocotrienols in hypercholesterolemic subjects. Functional Foods in Health and Disease. 2011;3:106-117.
- Ajuluchukwu JN, Okubadejo NU, Mabayoje M, Ojini FI, Okwudiafor RN, Mbakwem AC, Fasanmade OA, Oke DA. Comparative study of the effect of tocotrienols and αtocopherol on fasting serum lipid profiles in patients with mild hypercholesterolemia: a preliminary report. Niger Postgrad Med J. 2007;14(1):30-33.
- Baliarsingh S, Beg ZH, Ahmad J. The therapeutic impacts of tocotrienols in type 2 diabetic patients with hyperlipidemia. Atherosclerosis. 2005;182:367-374.
- Berger A, Rein D, Schafer A, Monnard I, Gremaud G, Lambelet P, Bertoli C. Similar cholesterol-lowering properties of rice bran oil, with varied γ-oryzanol, in mildly hypercholesterolemic men. Eur J Nutr. 2005;44:163-172.
- Qureshi AA, Sami SA, Salser WA, Khan FA. Dose-dependent suppression of serum cholesterol by tocotrienol-rich fraction (TRF₂₅) of rice bran in hypercholesterolemic humans. Atherosclerosis. 2002;161: 199-207.
- Qureshi AA, Sami SA, Salser WA, Khan FA. Synergistic effect of tocotrienol-rich fraction (TRF₂₅) of rice bran and lovastatin on lipid parameters in hypercholesterolemic humans. J Nutr Biochem. 2001;12:318-329.
- Qureshi AA, Bradlow BA, Salser WA, Brace LD. Novel tocotrienols of rice bran modulate cardiovascular disease risk parameters of hypercholesterolemic humans. J Nutr Biochem. 1997;8:290-298.
- Tomeo AC, Geller M, Watkins TR, Gapor A. Antioxidant effects of tocotrienols in patients with hyperlipidemia and carotid stenosis. Lipids. 1995;30(12):1179-1183.
- Qureshi AA, Qureshi N, Wright JJK, Shen Z, Kramer G, Gapor A, Chong YH, DeWitt G, Ong ASH, Peterson DM, Bradlow BA.

- Lowering of serum cholesterol in hypercholesterolemic humans by tocotrienols (palmvitee). Am J Clin Nutr. 1991;53:1021S-1026S.
- Tan DTS, Khor HT, Low WHS, Ali A, Gapor A. The effect of palm oil vitamin E concentrate on the serum and lipoprotein lipids in humans. Am J Clin Nutr. 1991;53: 1026S-1030S.
- Rasool AH, Yuen KH, Yusoff K, Wong AR, Rahman AR. Dose dependent elevation of plasma tocotrienol levels and its effect on arterial compliance, plasma total antioxidant status, and lipid profile in healthy humans supplemented with tocotrienol rich vitamin E. J Nutr Sci Vitaminol (Tokyo). 2006;52(6):473-478.
- 14. Mustad VA, Smith CA, Ruey PP, Edens NK, DeMickele SJ. Supplementation with 3 compositionally different tocotrienol supplements does not improve cardiovascular disease risk factors in men and women with hypercholesterolemia. Am J Clin Nutr. 2002;76:1237-1243.
- O'Byrne D, Grundy S, Packer L, Devaraj S, Baldenius K, Hoppe PP, Kreamer K, Jialal I, Traber MG. Studies of LDL oxidation following α-, γ-, or δ-tocotrienyl acetate supplementation of hypercholesterolemic humans. Free Radical Biol. Med. 2000;29(9):834-845.
- Mensink RP, Van-Houwelingen AC, Kromhout D, Hornstra GA: Vitamin E concentrate rich in tocotrienols had no effect on serum lipids, lipoproteins, or platelet function in men with mildly elevated serum lipid concentrations. Am J Clin Nutr. 1999;69(2):213-219.
- Wahlqvist ML, Krivokuca-Bogetic Z, Lo CH, Hage B, Smith R, Lukito W: Differential responses to tocopherols and tocotrienols during vitamin E supplementation in hypercholesterolemic individuals without change in coronary risk factors. Nutr Res. 1992;12:S181-S201.
- Khor HT, Chieng DY, Ong KK: Tocotrienols inhibit HMG-Co A reductase activity in the guinea pig. Nutr Res. 1995;15;537-544.
- Khor HT, Ng TT. Effects of administration of alpha-tocopherol and tocotrienols on serum lipids and liver HMG-Co A reductase activity. Int J Food Sci Nutr. 2000;51(Suppl):S3-11.

- Watkins T, Lenz P, Gapor AT, Struck M, Tomeo A, Bierenbaum M. γ-tocotrienol as a hypocholesterolemic and antioxidant agents fed atherogenic diets. Lipids. 1993;28:1113-1118.
- Qureshi AA, Tan X, Reis JC, Badr MZ, Papasian CJ, Morrison DC, Qureshi N. Suppression of nitric oxide induction and pro-inflammatory cytokines by novel proteasome inhibitors in various experimental models. Lipids in Health and Disease. 2011;10:177.
- Qureshi AA, Mo H, Packer L, Peterson DM. Isolation and identification of novel tocotrienols from rice bran with hypercholesterolemic, antioxidant, and antitumor properties. J Agri & Food Chemistry. 2000;48(8):3130-3140.
- Qureshi AA, Reis JC, Papasian CJ, Morrison DC, Qureshi N. Tocotrienols inhibit lipopolysaccharide pro-inflammatory cytokines in macrophages of female mice. Lipids in Health and Disease. 2010;9:143.
- Sen CK, Khanna S, Roy S. Tocotrienols in health and disease: The other half of the natural vitamin E family. Molecular Aspect of Medicine. 2007;28:692-728.
- Popa C, Netea MG, van Riel PL, van der Meer JW, Stalenhoef AF. The role of TNFalpha in chronic inflammatory conditions, intermediary metabolism, and cardiovascular risk. J Lipid Res. 2007;48(4):751-762.
- Simon AD, Yasdani S, Wand W, Schwartz A, Rabbani LE. Elevated plasma levels of interleukin-2 and soluble IL-2 receptor in ischemic heart disease. Clin Cardiol. 2001;24(3):253-256.
- Rosello-Lleti E, Rivera M, Bertomeu V, Cortes R, Jordan A, Gonzalez-Molina A. Interleukin-4 and cardiac fibrosis in patients with heart failure. Rev Esp Cardiol (Engl Ed). 2007;60(7):777-780.
- Kanda T, Takahashi T. Interleukin-6 and cardiovascular diseases. Japanese heart journal. 2004;45(2):183-193.
- 29. Apostolakis S, Vogiatzi K, Amanatidou V, Spandidos DA. Interleukin-8 and cardiovascular disease. Cardiovascular Research. 2009;84(3):353-360.
- 30. Small EM, Frost RJ, Olson EN. MicroRNAs add a new dimension to cardiovascular

- disease. Circulation. 2010;121(8):1022-1032.
- 31. Novak J, Kruzliak P, Bienertova-Vasku J, Slaby O, Novak M. MicroRNA-206: a promising theranostic marker. Theranostics. 2014;4(2):119-133.
- National Cholesterol Education Program. ATP III at a glance quick reference. NIH Publication. 2001;NO:01-3305.
- Kostner GM. Enzymatic determination of cholesterol in high-density lipoprotein fractions prepared by polyanion precipitation. Clin Chem. 1976;22:695-698.
- 34. Abacus Concepts, StatView Abacus Concepts, Inc., Berkeley, CA; 1992.
- Fenteany G, Standaert RF, Reichard GA, Corey EJ. A β-lactone related to lactacystin induces neurite outgrowth in a neuroblastoma cell line and inhibits cell cycle progression in an osteosarcoma cell line. Proc Natl Acad Sci. USA. 1994;91:3358-3362.
- 36. Lin KI, Baraban JM, Ratan RR. Inhibition versus induction of apoptosis by proteasome inhibitors depends on concentration. Cell Death and Differentiation, 1998:5:577-583.
- Schwarz K, Giuli RD, Schmidtke G, Kosstka S, Broek MVD, Kim KB, Crews CM, Kraft R, Groettrup M. The selective proteasome inhibitors lactacystin and epoxomicin can be used to either up- or down-regulate antigen presentation of nontoxic doses. J Immunology. 2000;164:6147-6157.
- Husain K, Centeno B, Perez M, Lee GZ, Sabiha K, Dung-Tsa C, Sebti S, Malafa M. Vitamin E delta-tocotrienol augments the antitumor activity of gemcitabine and suppresses constitutive NF-kappaB activation in pancreatic cancer. Mol Cancer Therapy. 2011;10(12):2363-2372.
- Qureshi AA, Pearce BC, Nor RM, Gapor AT, Peterson DM, Elson CE. α-Tocopherol attenuates the impact of γ-tocotrienol on hepatic 3-hudroxy-3-methylglutaryl coenzyme A reductase activity in chickens. J Nutr. 1996;126(2):389-394.
- Klein EA, Thomson IM Jr., Crowley JJ, Lucia MS, Goodman PJ, Minasian LM, Ford LG, Parnes HL, Gaziano JM, Karp DD, Lieber MM, Walther PJ, Klotz L, Parson JK, Chin JL, Darke AM, Lippman

- SM, Goodman GE, Meyskens FL Jr., Baker LH. Vitamin E and the risk of prostate cancer: the Selenium and Vitamin E Cancer Prevention Trial (SELECT). J Am Med Asso. 2011;306(14):1549-1556.
- 41. Freeman MR, Solomon KR. Cholesterol and benign prostate disease. Differentiation. 2011;82(4-5):244-252.
- Kok DEG, Roermund JGH van, Aben KKH, Heijer M den, Swinkels DW, Kampman E, Keimeney LALM. Blood lipid levels and prostate cancer risk; a cohort study. Prostate Cancer and Prostatic Diseases. 2011;14(4):340-345.
- 43. Shafique K, McLoone P, Qureshi K, Leung H, Hart C, Morrison DS. Cholesterol and the risk of grade-specific prostate cancer incidence: evidence from two large prospective cohort studies with up to 37 years' follow up. BioMedical Central Cancer. 2012;12:25.
- Murtola TJ, Syvala H, Pennanen P, Blauer M, Solakivi T, Ylikomi T, Tammela TJ. The importance of LDL and cholesterol metabolism for prostate epithelial cell growth. PLoS One. 2012;7(6):e39445.
- 45. Krycer JR, Phan L, Brown AJ. A key regulator of cholesterol homeostasis, SREBP-2, can be targeted in prostate cancer cells with natural products. Biochem J. 2012;446:191-201.
- Ding C, Cicuttini F, Li J, Jones G. Targeting IL-6 in the treatment of inflammatory and autoimmune diseases. Expert opinion on investigational drugs. 2009;18(10):1457-1466.
- Yam ML, Abdul Hafid SR, Cheng HM, Nesaretnam K. Tocotrienols suppress pro-inflammatory markers and cyclooxygenase-2 expression in RAW 264.7 macrophages. Lipids. 2009;44(9): 787-797.
- Ren Z, Pae M, Dao MC, Smith D, Meydani SN, Wu D. Dietary supplementation with tocotrienols enhances immune function in C57BL/6 mice. J Nutr. 2010;140(7):1335-1341.
- Khan DA, Ansari WM, Khan FA. Pro/antiinflammatory pathogenesis of premature coronary artery disease. J Interferon & Cytokines Research. 2011;31(7):561-567.
- 50. Menghini R, Stohr R, Federici M. MicroRNAs in vascular ageing and

- atherosclerosis. Ageing Research Reviews. 2014;03:005; DOI:10.1016/J. arr.
- Xu J, Zhao J, Evan G, Xiao C, Cheng Y, Xiao J. Circulating microRNAs: novel biomarkers for cardiovascular diseases. J Mol Med. 2012;90:865-875.
- 52. Fichtlscherer S, De Rosa S, Fox H, Schwietz T, Fischer A, Liebetrau C, Weber M, Hamm CW, Roxe T, Muller-Ardogan M, Bonauer A, Zeiher AM, Dmmeler S. Circulating microRNAs in patients with coronary artery disease. Circ Res. 2010;107:677-684.
- 53. Ono K, Kuwabara Y, Han J. MicroRNAs in cardiovascular diseases. Federation of European Biochemical Societies Journal. 2011;278(10):1619-1633.
- 54. Yan H, Wu J, Liu W, Zuo, Y, Chen, S, Zhang, S, Zeng, M, Huang, W: MicroRNA-20a overexpression inhibited proliferation and metastasis of pancreatic carcinoma cells. Human Gene Therapy. 2010;21(12): 1723-1734.
- 55. Shehadeh LA, Sharma S, Pessanha M, Wei JQ, Liu J, Yuan H, Rodrigues CO, Scherr M, Tsinoremas NF, Bishopric NH. MicroRNA-20a constrains p300-driven myocardial angiogenic transcription by direct targeting of p300. PloS One. 2013;8(11):e79133.
- Williams AH, Valdez G, Moresi V, Qi X, McAnally J, Elliott JL, Bassel-Duby R, Sanes JR, Olson EN. MicroRNA-206 delays ALS progression and promotes regeneration of neuromuscular synapses in mice. Science. 2009;326(59):1549-1554.
- 57. Ren J, Huang HJ, Gong Y, Yue S, Tang LM, Cheng SY. MicroRNA-206 suppresses gastric cancer cell growth and metastasis. Cell & Bioscience. 2014;4:26.
- Kamisah Y, Qodriyah HM, Chua KH, Nur Azlina MF. Vitamin E: A potential therapy for gastric mucosal injury. Pharmaceutical Biology. 2014;1-7.
- McNally EM, Goldstein JA. Interplay between heart and skeletal muscle disease in heart failure: the 2011 George E. Brown Memorial Lecture. Circ Res. 2012;110(5): 749-754.
- Limana F, Esposito G, D'Arcangelo D, Di Carlo A, Romani S, Melillo G, Mangoni A, Bertolami C, Pompilio G, Germani A, Capogrossi MC. HMGB1 attenuates

- cardiac remodelling in the failing heart via enhanced cardiac regeneration and miR-206-mediated inhibition of TIMP-3. PloS One. 2011;6(6):e19845.
- 61. Das S, Mukherjee S, Lekli I, Gurusamy N, Bardhan J, Rachoudhury U, Chakravarty
- R, Banerji S, Knowlton AA, Das DK. Tocotrienols confer resistance to ischemia in hypercholesterolemic hearts: insight with genomics. Mol Cell Biochem. 2012;360:35-45.

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