

## Gelatin chitosan film incorporated with clove essential oil for retaining quality of silver pomfret fish fillet

Aidilla Mubarak<sup>1\*</sup>, Zakirah Safi Othman<sup>1</sup>, Nurul Ulfah Karim<sup>2</sup>

<sup>1</sup>School of Food Science and Technology, Universiti Malaysia Terengganu, 21030 Kuala Nerus, Terengganu, Malaysia

<sup>2</sup>School School of Fisheries and Aquaculture Science, Universiti Malaysia Terengganu, 21030 Kuala Nerus, Terengganu, Malaysia

Received:  
September 10, 2018

Accepted:  
February 01, 2019

Published:  
March 30, 2019

### Abstract

Clove (*Syzygium aromaticum* L.) essential oil has been reported for its potent antioxidant and antimicrobial activities. In this study, clove essential oil was incorporated with gelatin-chitosan solution to develop an edible film (CEO film) for fish preservation. The objective was to determine the effectiveness of this edible film in controlling physical, biochemical and microbial changes in fillet of silver pomfret (*Pampus argenteus*). The effectiveness of this film was compared with uncoated fillet (control), and gelatin-chitosan film (GC film). The formulated film was tested for water solubility and antimicrobial activity against four selected microorganisms: *Escherichia coli*, *Pseudomonas aeruginosa*, *Salmonella enterica* and *Bacillus cereus*. The effectiveness of the formulated film on the silver pomfret fillet was evaluated based on weight loss, pH, firmness, total volatile basic nitrogen (TVB-N) and total plate count (TPC) of the fish fillet. The CEO films intermediately inhibit the growth of *E. coli* and *S. enterica*. Meanwhile, GC film did not show inhibition on the growth of tested microorganisms. CEO film had lower water solubility compared to GC film. CEO film was observed to reduce weight loss ( $p < 0.05$ ), lower pH on day 6 ( $p < 0.05$ ), and increase firmness of fish fillet when compared to the control ( $p < 0.05$ ). Fish fillet applied with CEO film also had lower TVB-N value and microbial count. This study shows that the CEO film has antimicrobial properties which can benefit fish preservation. Improvement for developing the edible film with acceptable properties is thus important to extend the shelf life of fish fillet.

**Keywords:** Clove, Essential oil, Edible film, Fish fillet, Shelf life

### How to cite this:

Mubarak A, Othman ZS and Karim NU, 2019. Gelatin chitosan film incorporated with clove essential oil for retaining quality of silver pomfret fish fillet. Asian J. Agric. Biol. 7(1): 86-92.

\*Corresponding author email:  
aidilla@umt.edu.my

This is an Open Access article distributed under the terms of the Creative Commons Attribution 3.0 License. (<https://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## Introduction

Silver pomfret (*Pampus argenteus*) is one of the important fish species with high market value (Zhao et al., 2010). The silver pomfret plays an important role in the fisheries sector of many countries in the world

including Malaysia, China, India, Thailand, Japan, Korea, Kuwait and Iran (FAO, 1995). Silver pomfret is highly perishable during post-mortem aging. Studies have explored various approaches on extending shelf life of fish and fishery products, including packaging techniques, edible film and utilization of low storage



temperature (Lakshmanan et al., 2002; Fan et al., 2009; DeWitt and Oliveira, 2016).

Spices have been largely utilized in food as flavoring agents as well as food preservatives and this is attributed to its antioxidant and antimicrobial activities. Clove (*Syzygium aromaticum* L.) in particular has been reported for its potent antioxidant and antimicrobial activities (Shan et al., 2005). Their applications as preservatives in food or antiseptics and disinfectants have been widely investigated. Essential oil fractions are found to contain active compounds that provide valuable antimicrobial activity, against food-borne pathogens and spoilage bacteria (Gutierrez et al., 2008). Clove essential oil which contains mainly eugenol was reported as an effective inhibitor of pathogenic bacteria in previous studies (Cressy et al., 2003; Mytle et al., 2006). Edible films incorporated with various essential oils were shown to have antimicrobial property against foodborne pathogens (Fernández-Pan et al., 2012).

This study aims to identify the effect of gelatin-chitosan films incorporated with clove essential oil in controlling physical, biochemical and microbial changes of silver pomfret fillet.

## Material and Methods

### Film formulation

Gelatin from bovine-hide in combination with chitosan (Guinama, Valencia) were used to develop the film forming solution as previously described in Gomez-Estaca et al. (2010). For gelatin-chitosan film forming solutions, 6 g of gelatin (dissolved in distilled water) and 1 g of chitosan (dissolved in 0.15 M acetic acid) per 100 ml of film forming solution were used. Sorbitol and glycerol (0.15g/g biopolymer, gelatin plus chitosan) were added as plasticizers, then warmed and stirred at 45°C for 15 min. In preparation of the clove-added film forming solution, food grade clove essential oil was added at the proportion of 0.75 ml/g biopolymer. Soya lecithin (0.125 g/g biopolymer) was added as an emulsifying agent. The clove-added film forming solution was homogenized with an Ultraturax blender (position 5, 1 min). 5ml of the solutions were casted in a petri dish and dried at 45°C in a forced-air oven for 15 h to yield a film with uniform thickness of 200 µm. Two different types of films were then obtained: a gelatin-chitosan film (GC) and a gelatin-chitosan clove-added film (CEO).

### Water solubility

Film portions measuring 1 cm<sup>2</sup> was placed in aluminum capsules with 15 ml of distilled water and gently shaken at 22°C for 15 h. The solution was then filtered using Whatman no. 1 filter paper to recover the remaining undissolved film. The undissolved film was then desiccated at 105°C for 24 h. Film solubility is calculated according to Gómez-Estaca et al. (2010) using the following equation:

$$\text{Film solubility, FS(\%)} = \left( \frac{W_0 - W_f}{W_0} \right) \times 100$$

Where

$W_0$  = initial weight of the film expressed as dry matter

$W_f$  = weight of the undissolved desiccated film residue

### Antimicrobial activity measurement

The formulated films were tested for antimicrobial activity over four microorganisms: *Escherichia coli*, *Pseudomonas aeruginosa*, *Bacillus cereus*, and *Salmonella enterica* according to Duan et al. (2010) with modifications. Spread plates of BHI Agar were inoculated with 100 µl of the respective bacteria and grown overnight (10<sup>8</sup> CFU/ml). Then, circular pieces of the each formulated film (1.5 cm in diameter) were laid onto the surface of the inoculated plate. After incubation, the observed inhibition zones-surrounding clear areas were measured for indication of its antimicrobial activity.

### Fish storage trial

Silver pomfret (*P. argenteus*) were cut in fillets (5 cm x 4 cm) and divided into three groups which are uncoated, coated with gelatin-chitosan film (GC) and coated with the gelatin-chitosan films containing clove essential oil (CEO). All fillets were then vacuum-packed in HDPE and stored at 2°C for 8 days. All treatments were done in triplicate. The uncoated and coated fillets were then measured for its physical, chemical and microbiological changes.

### Weight loss of fish fillet

The weight of uncoated, GC and CEO fillets were aseptically measured at an interval of two days until the end of storage period. The weight loss were then calculated and presented as percentage of weight loss.

### pH of fish fillet

Approximately 5-10 g of fish fillet was homogenized with a double quantity of distilled water. After 5 min at ambient temperature, the pH was determined. pH



value was measured at an interval of two days until the end of storage period.

### Firmness of fish fillet

Textural properties of the fish fillet were assessed using the puncture test with a TA-XT2 Texture Analyser. The maximum force required to puncture the fish fillets or films was recorded. Compression applied using P2N (needle) at a speed of 1.0 mms<sup>-1</sup> was used for puncturing the plugs position over a 3-mm hole. Three replicates were analysed for each treatment. Firmness of all fillets was measured at an interval of two days until the end of storage period.

### Total volatile basic nitrogen (TVB-N) of fish fillet

Total volatile basic nitrogen (TVB-N) was assessed according to Cobb III et al. (1973) with some modification. Fish fillet (1 g) was loaded into Kjeldahl-type distillation tube and added with 12 ml of concentrated sulphuric acid for protein digestion along with one tab of Kjeltabs Cu-3.5. After digestion, 40% sodium hydroxide (NaOH) solution (50 ml) was added and followed by 75 ml of distilled water. The distillation was performed and the distillate was collected into a conical flask filled with 3% boric acid solution and methyl red and bromocresol green indicator. The solution was then titrated with 0.05 N hydrochloric acid (HCl). TVB-N was presented as nitrogen per 100 g calculated with the following equation:

$$\text{Nitrogen} = \frac{\text{titre volume} \times 0.05 \times 14}{\text{weight of sample in grams}}$$

TVB-N of fillet for uncoated, GC and CEO coated fillet were determined at the last two days of the storage period.

### Microbiological count of fish fillet

Microbial count was determined through total plate count. A total amount of 10 g of fish fillet were placed in a sterile plastic bag with 90 ml of buffered 0.1% peptone water in a vertical laminar-flow cabinet. Appropriate dilutions were prepared and spread onto PCA agar, which was then incubated at 35°C for 24 h. All microbiological counts were expressed as the log of the colony-forming units per gram (log CFU/g) of sample. The analysis was performed at the end of the storage period. Analysis was done in triplicates.

### Statistical analysis

Data analysis was carried out using SPSS version 20.0. The differences of mean among treatment were analyzed using one-way analysis of variance (ANOVA) and Tukey test, with a level of significance at  $p < 0.05$ .

## Results and Discussion

### Water solubility of formulated fish film

Figure 1 shows the solubility of the two formulated edible films (gelatin-chitosan film, GC and gelatin-chitosan incorporated with clove essential oil, CEO). GC film showed a higher solubility value of (76.5%) than the CEO film (8.98%). This indicated that the CEO film is better in terms of potential protection for the fish fillet. Edible film with low solubility enhances product integrity and its water resistance (Perez-Gago and Krochta, 1999). Films solubility is often designed to be of low solubility to aid in protecting product from humidity and water loss (Gontard et al., 1993). The conjoint hydrophobic property of oil and gelatin could enhance the hydrophobicity of our CEO formulated film (Ahmad et al., 2012).

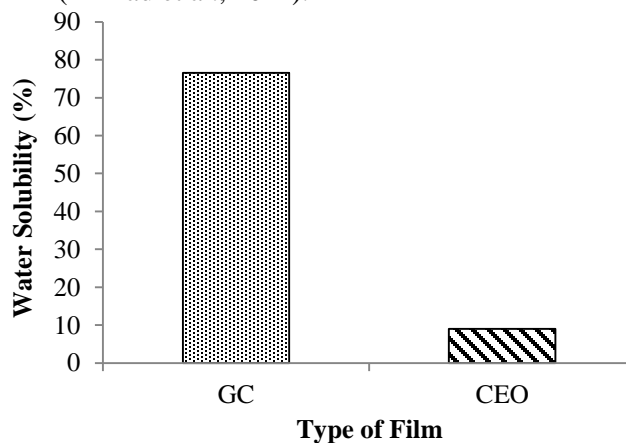


Fig. 1. Water solubility of formulated film

### Antimicrobial activity of formulated fish film

Table 1 summarizes the results from antimicrobial test of formulated film against selected microorganism. GC film did not show any inhibition on growth of selected microorganism (*Escherichia coli* 2922, *Pseudomonas aeruginosa*, *Bacillus cereus*, and *Salmonella enterica*). CEO film showed an intermediate inhibition against growth of *E.coli* 2922 and *S. enterica*. Components of clove oil were reported to have efficiency in inhibiting growth of microorganisms (Burt, 2004). Clove oil has chemical

compositions characterized by terpenoids (eugenol) which are active against a broad range of microorganism (Dorman and Deans, 2000). Eugenol was reported to suppress the growth of *L.monocytogenes*, *C. jejuni*, *S. Enteritidis*, *E. coli* and *S. aureus* (Cressy et al., 2003).

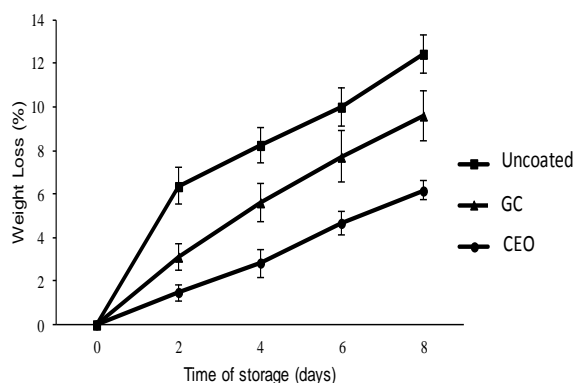
**Table 1. Antimicrobial activity of formulated film**

Type of film	<i>Escherichia coli</i>	<i>Bacillus cereus</i>	<i>Salmonella enterica blue</i>	<i>Pseudomonas aeruginosa</i>
GC	X	X	X	X
CEO	✓	X	✓	X

\* ✓ : inhibition, x: no inhibition zone

**Effect of formulated films on weight loss of fish fillet**

Figure 2 illustrates the percentage of weight loss in uncoated, GC and CEO coated fillet. Both GC and CEO coated fillet had a significantly reduced percentage of weight loss when compared to uncoated fillet ( $p < 0.05$ ). Moreover, addition of clove oil (CEO) showed a tendency for a lesser weight loss in the fish fillet when compared to GC, but there was no significant difference among the two treatment groups.



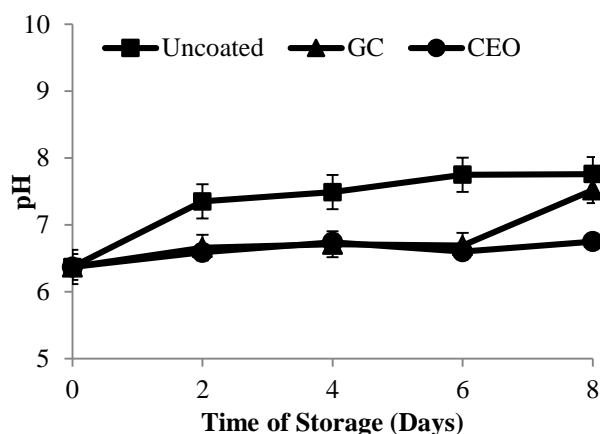
**Fig. 2. Effect of different treatments on weight loss of silver pomfret fish fillet (data are presented as mean ± standard deviation)**

Apart from storage temperature, postharvest treatment is known to influence the rate of weight loss (Hernández-Muñoz et al., 2006). Coating was previously reported to be effective in reducing water loss and in prolonging shelf life of fish fillets (Jeon et

al., 2002). Incorporation of essential oil into coating such as chitosan-oregano oil coating was reported to be effective in lowering weight loss in fruit (Wu et al., 2016) which supported our observation in this study. Oregano oil with combination of oxygen absorber was also found to reduce weight loss in rainbow trout (Mexis et al., 2009). Coating may act as an effective barrier to oxygen, carbon dioxide and water vapour transmission which helps reduce the loss of weight in produce (de Aquino et al., 2015).

**Effect of formulated films on pH of fish fillet**

Figure 3 shows the effect of different treatments (uncoated, GC and CEO coating) on the pH of the fish fillets. There was a tendency for lower pH value in fillet coated with GC and CEO when compared to uncoated fillet from day 2 and 4 after treatment, but no significant difference were detected between the treatment groups. On day 6 and 8 after treatment, the CEO coated had a significantly lower pH value when compared to the uncoated fillet ( $p < 0.05$ ). The increase of pH could be a result of accumulation of alkaline compounds which includes ammonia and trimethylamine which are produced during spoilage of fish (Ruiz-Capillas and Moral, 2005; Özyurt et al., 2009). Therefore, the decrease in pH of CEO film coated fillet indicated an inhibition of fish spoilage.



**Fig. 3. Effect of different treatments on pH of silver pomfret fish fillet (data are presented as mean ± standard deviation)**

**Effect of formulated films on texture changes of fish fillet**

Figure 4 shows the effect of different treatments (uncoated, GC and CEO coating) on the firmness of the fish fillets. No significant difference was observed



between treatments, although there was a slight tendency for a slower reduction in firmness of CEO coated compared to uncoated fish fillet.

Autolytic enzymes reduces textural quality during early stages of deterioration when spoilage characteristics such as off-odors and off-flavors are not yet prevalent (Hansen et al., 1996). Microbial action, proteolytic activity and water loss in the muscle could result in degradation in protein which thereafter affects the texture loss in the fish meat (Suárez-Mahecha et al., 2007; Pacheco-Aguilar et al., 2008). Softening of the fish meat is a critical quality deterioration of fish meat that would largely affect consumer's acceptance.

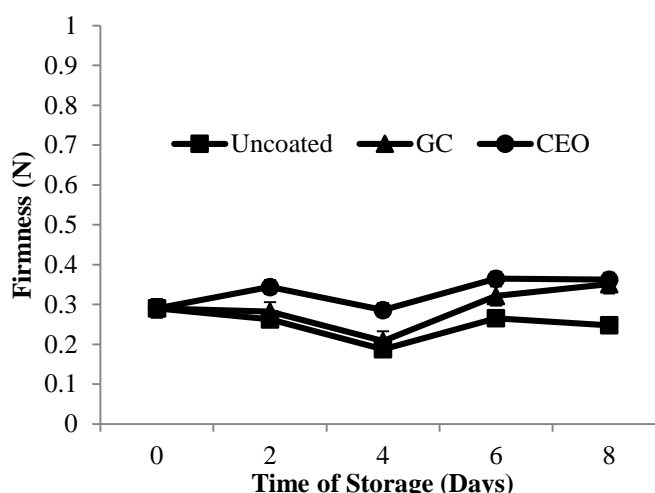


Fig. 4. Effects of different treatments on firmness of silver pomfret fish fillet (data are presented as mean ± standard deviation)

**Effect of formulated films on TVB-N content of fish fillet**

Table 2 shows the increase of TVB-N content in fish fillet during the last 2 days of storage with different treatments (uncoated, GC, CEO coating). CEO coated fish fillet showed the lowest TVB-N increase when compared to GC and uncoated fish fillet (p<0.05). Rotting fish has an enhanced level of TVB-N specifically its major constituents which are ammonia, dimethylamine and trimethylamine. During fish spoilage, bacterial deamination and autolytic breakdown of adenosine monophosphate occur which produces ammonia (Shakila et al., 2003). Antibacterial properties of clove essential oil may influence the lower TVB-N content in samples treated with CEO (Burt, 2004).

Ojagh et al. (2010) reported similar results on the effect of chitosan coating added with essential oil on rainbow trout meat.

**Table 2. Increase of TVB-N content in silver pomfret fish fillet during the last 2 days of storage with different treatments**

Treatments	Differences in TVB-N value from day 6 to day 8 (mg/100g)
Uncoated	4.2
GC	1.07
CEO	0.45

**Effect of formulated films on total plate count of fish fillet**

Table 3 shows the effect of different treatments (uncoated, GC, CEO coating) on total plate count of fish fillets. The result indicated a reduced microbial growth in fillet coated with CEO film compared to the GC film and uncoated fillet. In a previous study, incorporation of essential oil into gelatin-chitosan film was reported to reduce the growth of microorganism in fishes (Gomez-Estaca et al., 2010). In another study, a coating made up of chitosan and gelatin also showed an inhibitory effect on the gram-negative microbes of fish flesh (L’opez-Caballero et al., 2005), which supports our finding.

**Table 3. Total plate count of silver pomfret fish fillet at the end of storage with different treatments**

Treatment	Average
	Day 8 (cfu/g)
Uncoated	TMTC
GC	TMTC
CEO	1.91 x 10 <sup>9</sup>

\* TMTC: too many to count

**Conclusion**

In conclusion, the addition of clove essential oil to formulation of gelatin-chitosan film allowed the attainment of edible films with antimicrobial properties and low water solubility. Gelatin-chitosan film incorporated with clove oil reduced the weight loss and the microbial growth of the fish fillet. The application of this edible film also delayed the increase of TVB-N value indicating a slower rate of spoilage. The effects provided by the formulated edible film can help in prolonging shelf life of the fish fillet.



## Acknowledgement

The authors acknowledge the support of funding from School of Food Science and Technology, Universiti Malaysia, Terengganu.

## Contribution of Authors

Mubarak A: Design of work, data analysis and interpretation, drafting and critical revision of the article and final approval of the version to be published.

Othman ZS: Data collection, data analysis, data interpretation and drafting of the article.

Karim NU: Data analysis, data interpretation and drafting of the article.

**Disclaimer:** None.

**Conflict of Interest:** None.

**Source of Funding:** School of Food Science and Technology, University Malaysia, Terengganu, Malaysia.

## References

- Ahmad M, Benjakul S, Prodpran T and Agustini TW, 2012. Physico-mechanical and antimicrobial properties of gelatin film from the skin of unicorn leatherjacket incorporated with essential oils. *Food Hydrocoll.* 28(1): 189-199.
- Burt S, 2004. Essential oils: Their antibacterial properties and potential applications in foods—A review. *Int. J Food Microbiol.* 94(3): 223–253.
- Cobb III GF, Alaniz I and Thompson Jr CA, 1973. Biochemical and microbial studies on shrimp: volatile nitrogen and amino nitrogen analysis. *J. Food Sc.* 38(3): 431-436.
- Cressy HK, Jerrett AR, Osborne CM. and Bremer PJ, 2003. A novel method for the reduction of numbers of *Listeria monocytogenes* cells by freezing in combination with an essential oil in bacteriological media. *J. Food Protect.* 66(3): 390-395.
- de Aquino AB, Blank AF and de Aquino Santana LCL, 2015. Impact of edible chitosan-cassava starch coatings enriched with *Lippia gracilis* Schaeur genotype mixtures on the shelf life of guavas (*Psidium guajava* L.) during storage at room temperature. *Food Chem.* 171: 108-116.
- DeWitt CAM and Oliveira ACM, 2016. Modified atmosphere systems and shelf life extension of fish and fishery products. *Foods.* 5(3): 48.
- Dorman HJD and Deans SG, 2000. Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *J. Appl. Microbiol.* 88(2): 308-316.
- Duan J, Cherian G, and Zhao Y, 2010. Quality enhancement in fish and frozen lingcod (*Ophiodon elongates*) fillets by employment of fish oil incorporated chitosan coatings. *Food Chem.* 119(2): 524-532.
- Fan W, Sun J, Chen Y, Qiu J, Zhang Y and Chi Y, 2009. Effects of chitosan coating on quality and shelf life of silver carp during frozen storage. *Food Chem.* 115(1): 66-70.
- FAO, 1995. Yearbook of Fishery Statistics. Catches and Landings. Volume 80. 302–306.
- Fernández-Pan I, Royo M and Maté JI, 2012. Antimicrobial activity of whey protein isolate edible films with essential oils against food spoilers and foodborne pathogens. *J. Food Sci.* 77(7): 383-390.
- Gómez-Estaca J, de Lacey López A, López-Caballero ME, Gómez-Guillén MC and Montero P, 2010. Biodegradable gelatin–chitosan films incorporated with essential oils as antimicrobial agents for fish preservation. *Food Microbiol.* 27(7): 889-896.
- Gontard N, Guilbert S. and Cuq JL, 1993. Water and glycerol as plasticizer affect mechanical and water vapor barrier properties of a wheat gluten film. *J. Food Sci.* 58: 206-211.
- Gutierrez J, Barry-Ryan C and Bourke P, 2008. The antimicrobial efficacy of plant essential oil combinations and interactions with food ingredients. *Int. J. Food Microbiol.* 124(1): 91-97.
- Hansen TL, Gill T, Rontved SD and Huss HH, 1996. Importance of autolysis and microbiological activity on quality of cold-smoked salmon. *Food Res. Int.* 29(2): 181-186.
- Hernández-Muñoz P, Almenar E, Ocio MJ and Gavara R, 2006. Effect of calcium dips and chitosan coatings on postharvest life of strawberries (*Fragaria × ananassa*). *Postharvest Biol. Technol.* 39(3): 247–253.
- Jeon YJ, Kamil JYVA and Shahidi F, 2002. Chitosan as edible invisible film for quality preservation of herring and Atlantic cod. *J. Agric. Food Chem.* 50(18): 5167–5178.
- Lakshmanan R, Shakila RJ and Jeyasekaran G, 2002. Survival of amine-forming bacteria during the ice storage of fish and shrimp. *Food Microbiol.* 19(6):



- 617-625.
- L'opez-Caballero ME, G'omez-Guill'en MC, P'erez-Mateos M and Montero P, 2005. A chitosan-gelatin blend as a coating for fish patties. *Food Hydrocoll.* 19(2): 303-311.
- Mexis SF, Chouliara E. and Kontominas MG, 2009. Combined effect of an oxygen absorber and oregano essential oil on shelf life extension of rainbow trout stored at 4°C. *Food Microbiol.* 26(6): 598-605.
- Mytle N, Anderson GL, Doyle MP and Smith MA, 2006. Antimicrobial activity of clove (*Syzygium aromaticum*) oil in inhibiting *Listeria monocytogenes* on chicken frankfurters. *Food Control*, 17(2): 102-107.
- Ojagh SM, Rezaei M, Razavi SH and Hosseini SMH, 2010. Effect of chitosan coatings enriched with cinnamon oil on the quality of refrigerated rainbow trout. *Food Chem.* 120(1):193-198.
- Özyurt G, Kuley E, Özkütük S and Özogul F, 2009. Sensory, microbiological and chemical assessment of the freshness of red mullet (*Mullus barbatus*) and goldband goatfish (*Upeneus moluccensis*) during storage in ice. *Food Chem.* 114(2): 505-510.
- Pacheco-Aguilar R, Márquez-Ríos E, Lugo-Sánchez ME, García-Sánchez G, Maeda-Martínez AN and Ocaño-Higuera VM, 2008. Postmortem changes in the adductor muscle of Pacific lions-paw scallop (*Nodipecten subnodosus*) during ice storage. *Food Chem.* 106(1): 253-259.
- Perez-Gago MB and Krochta JM, 1999. Water vapor permeability, solubility and tensile properties of heat-denatured versus native whey protein films. *J Food Sci.* 64(6): 1034-1037.
- Ruiz-Capillas C and Moral A, 2005. Sensory and biochemical aspects of quality of whole bigeye tuna (*Thunnus obesus*) during bulk storage in controlled atmospheres. *Food Chem.* 89(3): 347-354.
- Shakila RJ, Vijayalakshmi K and Jeyasekaran G, 2003. Changes in histamine and volatile amines in six commercially important species of fish of the Thoothukkudi coast of Tamil Nadu, India stored at ambient temperature. *Food Chem.* 82(3): 347-352.
- Shan B, Cai YZ, Sun M and Corke H, 2005. Antioxidant capacity of 26 spice extracts and characterization of their phenolic constituents. *J. Agric. Food Chem.* 53(20): 7749-7759.
- Suárez-Mahecha H, De Francisco A, Beirão LH, Pardo-Carrasco S and Córtes-Rodríguez M, 2007. Post mortem loss of texture of fish meat during cold storage. *Acta Biolo Colomb.* 12(1): 3-18.
- Wu C, Fu S, Xiang Y, Yuan C, Hu Y, Chen S, Liu D and Ye X, 2016. Effect of chitosan gallate coating on the quality maintenance of refrigerated (4°C) Silver Pomfret (*Pampus argenteus*). *Food Bioproc Tech.* 9(11): 1835-1843.
- Zhao F, Zhuang P, Zhang L and Shi Z, 2010. Biochemical composition of juvenile cultured vs. wild silver pomfret, *Pampus argenteus*: determining the diet for cultured fish. *Fish Physiol. Biochem.* 36(4): 1105-1111.